

Research



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Author for correspondence:

Reuven Dukas

e-mail: dukas@mcmaster.ca

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Animal behaviour

Genetic correlation between aggressive signals and fighting

Carling M. Baxter¹, Ieta Shams¹, Ian Dworkin² and Reuven Dukas¹

¹Department of Psychology, Neuroscience & Behaviour, and ²Department of Biology, McMaster University, 1280 Main Street West, Hamilton, Ontario, Canada L8S 4K1

ID, 0000-0002-2874-287X; RD, 0000-0003-4533-8542

Theoretical analyses indicate that aggressive signals should positively correlate with the signallers' willingness and abilities to fight. Few experimental studies, however, have tested this prediction. In two experiments employing distinct, ecologically realistic protocols, we quantified the association between aggressive signals and fighting in fruit fly genotypes and found high positive genetic correlations between threat and fighting ($r_G = 0.80$ and 0.74). Our results add to the growing body of experimental work indicating that aggressive signals have relatively high informational value.

1. Introduction

Many animals rely on aggression for access to desirable resources and mates. While such aggression has obvious potential fitness advantages, fighting may also have severe costs, which range from lost time and energy to wounding and death [1–3]. The vast majority of aggressive encounters, however, do not involve fighting. Rather, contenders most often settle confrontations by performing threat displays, which typically lead to one of the participants retreating [4–6].

Early theoretical analyses readily recognized the difficulty with threat displays: it pays all individuals including weak ones to threaten others if threat leads to the challenger withdrawing [7]. The only solution to this theoretical dilemma was to assume that threat either allows fair assessment of fighting ability or indicates willingness to escalate into fighting. In the latter case, contenders must use threat reliably in order to avoid costly loss against stronger opponents. This implies a high positive correlation between threat displays and fighting abilities [8–10].

While the theoretical challenge may have been resolved, the models on aggressive signals and fighting inspired critical reassessment [11] as well as new experiments designed to quantify the information conveyed by aggressive signals. Some studies examined whether threat displays signalled tendencies to escalate into fighting [12,13]. Most notably, in a few species of song birds, soft song was significantly associated with subsequent attack [14–17]. We know, however, of no studies that have examined the genetic correlation between threat and fighting.

A complementary approach for predicting the strength of association between threat signals and fighting is to consider likely mechanistic constraints. One would expect similar genetic, physiological and neuronal mechanisms to mediate aggressive signals and actual fighting. This could lead to a positive genetic correlation between threat and combat. Indeed both phenotypic and genetic correlations among traits are ubiquitous [18–22]. We lack data, however, about the genetic correlation between threat and fighting.

To further our understanding of the association between threat displays and fighting, we used fruit flies (*Drosophila melanogaster*), which are a highly tractable model system for quantifying aggression. When placed in settings that allow resource defence, male fruit flies defend attractive food patches both in

the absence and presence of females. The flies perform well-characterized aggressive behaviours. The sole, prominent aggressive signal is wing threat, in which a fly raises his wings at 45° toward his opponent. Fighting primarily includes lunging, where the aggressor hits his opponent with his forelegs, and occasionally more escalated combat including boxing, where both males rear up on their hind legs and strike each other with their forelegs; and tussling, involving the males stumbling over each other [23–27].

One of the unique tools available in fruit flies is the *Drosophila* Genetic Reference Panel (DGRP), a set of sequenced, fully inbred lines, which has been used widely for investigating the genetic basis of various traits including aggression [28–33]. We thus used a subset of the DGRP lines to test the genetic correlation between threat and fighting. Both signals of aggression and fighting itself may vary as a function of the social context [26,34]. Hence we conducted two experiments, each assessing threat and fighting in one of two realistic social settings. In natural sites with small fruits and low to moderate fly densities, capable males attempt to monopolize fruit through aggression and mate with females that seek food and egg laying sites. Sometimes, males and females co-occur at the fruits, while at other times, males may encounter only other males at the fruits [27]. We simulated these two relevant scenarios by allowing males in one experiment to interact with both males and females but to encounter only males in the other experiment. In both cases, we predicted a positive genetic correlation between threat displays and fighting.

2. Methods

(a) General

We used *Wolbachia*-free lines from the DGRP [33,35] and housed them in standard fly vials with 5 ml of our standard food medium (1 l = 90 g sucrose, 75 g cornmeal, 10 g carrageenan, 32 g yeast and 2 g methyl paraben dissolved in 20 ml ethanol). We maintained the flies in an environmental chamber at 25°C and 50% relative humidity on a 12 h light : dark cycle with the lights turning on at 10.00 h. In order to lessen deleterious inbreeding effects, we created F1 hybrid flies for the experiments by crossing males from 24 distinct DGRP lines to females from a single line (DGRP-83) [36]. To generate the hybrids, we collected virgin females of DGRP-83 within 8 h of eclosion using light CO₂ anaesthesia and housed them in groups of 15 per food vial sprinkled with live yeast to stimulate egg laying. Once females were 3–5 days old, we transferred them in groups of eight to new vials containing food and live yeast and added to each vial five males of 1–4 days old from one of the DGRP lines. We transferred these parental flies of the 24 hybrid crosses into new food vials with live yeast daily, and scraped excess eggs from the vials to ensure a consistent rearing density across lines. Eleven days after egg laying, we collected by aspiration the F1 hybrid flies (hereafter genotypes) within 8 h of eclosion to ensure virginity and aspirated the focal flies individually into food vials.

(b) Aggression experiments

We conducted two experiments to assess threat and fighting under the two realistic settings of males at a food patch that either has or lacks females. We tested the focal flies when they were 4 days old within 2 h of the onset of the light period. This period is associated with the morning peak of fly activity in most studies [27,37]. Our test arenas consisted of polystyrene Petri dishes that were 35 mm in diameter and 8 mm high. To

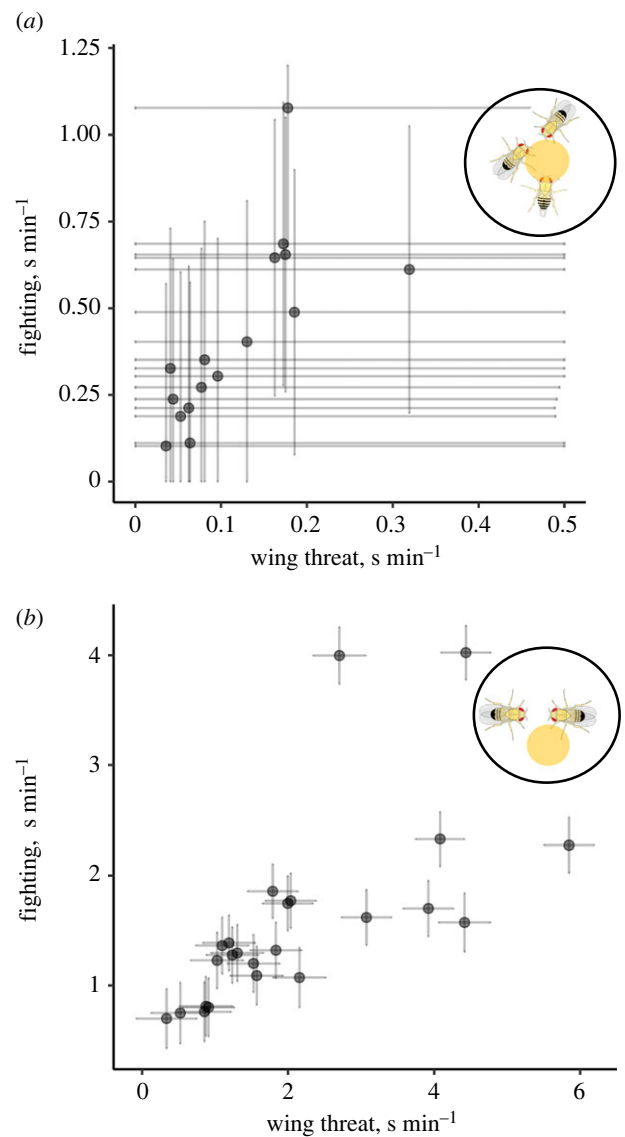


Figure 1. The genetic correlation between wing threat and fighting among distinct genotypes in the presence (a) and absence (b) of females. Note the different trait value ranges between (a) and (b). Each point shows conditional genotypic means \pm 95% CI. The inserts illustrate the arenas with and without females. Males are identified by the black tip of their abdomen. $N = 443$ trials with 16 genotypes and 361 trials with 24 genotypes, respectively.

restrict the flies to the floors of the arenas, we coated the walls and ceilings with Surfasil (Thermo Fisher, Ottawa, Ontario, Canada). We covered the floor of each arena with filter paper and placed at its centre a small, circular food patch made of standard medium. In experiment 1 (females present), the food patch was 7.5 mm in diameter and 1.5 mm high and coated with a suspension made of 3 g live yeast and 100 ml grapefruit juice. In experiment 2 (females absent), the food patch was 5 mm in diameter and had at its centre a 3 mm ball of paste made from a mixture of 5 g live yeast and 10 ml grapefruit juice. On each test day, we ran 1–4 sets of test trials for each of the genotypes with the order of genotypes randomized and counterbalanced. Experiment 1 included 16 DGRP genotypes and 443 trials (25–30 replicates per genotype). In experiment 2, we increased the number of DGRP genotypes to 24 and had a total of 361 trials (14–16 replicates per genotype).

In experiment 1 (females present), we aspirated into each arena two focal males and a recently mated female. Within each test arena, we always placed flies of a single genotype that had developed in distinct food vials to avoid familiarity. The female had mated once with a same-genotype male just

Table 1. Trait means (response scale), genetic variances, residual variances and heritabilities (latent variable scale).

experiment	behaviour	trait mean (95% CI)	V_G (95% CI)	V_r (95% CI)	H^2
females present	threats	0.093 (0.033,0.26)	1.30 (0.50, 3.22)	3.3 (2.4, 4.4)	0.28
females present	fighting	0.34 (0.12, 0.95)	1.30 (0.52, 3.28)	3.4 (2.7, 4.4)	0.27
females absent	threats	1.64 (0.98, 2.77)	1.24 (0.64, 2.50)	1.4 (1.0, 1.9)	0.47
females absent	fighting	1.4 (0.89, 2.19)	0.56 (0.44, 1.12)	0.64 (0.46, 0.90)	0.46

prior to the test. Recently mated females typically do not remate, and we excluded only two trials owing to mating. Female presence, however, may alter the dynamics of male aggression [26,34,38]. In experiment 2 (females absent), we aspirated into each arena just two focal males of the same genotype. We allowed the flies 5 min to acclimate and then video recorded them for 10 min using webcams (Logitech HD Pro C920). After the experiments, observers blind to fly genotype recorded from the videos the duration of wing threat and fighting [23,26] using BORIS, an event-logging software [39]. We recorded wing threat, which consists of the aggressor raising his wings at 45° toward his opponent. For fighting, we recorded lunging, which is the most common component of physical aggression and characterized by the aggressor making a fast movement and hitting with his head his opponent. We also recorded less frequent elements of fighting including holding, in which the aggressor uses his forelegs to grasp his opponent; boxing, where both males rear up on their hind legs and strike each other with their forelegs; and tussling, involving the males stumbling over each other (see [27], electronic supplementary material video S1 0:51). In the females-present experiment, males spent some time pursuing the females, which, theoretically, could reduce their time available for aggression. We nevertheless reported the total time males devoted to threat and fighting in order to avoid potential biases.

(c) Statistics

We analysed the data with R v. 4.0.2 [40]. We used tidyverse v. 1.3.2 [41] to organize data and generate figures, and constructed generalized linear mixed-effects models (Tweedie distributed with a log link) using glmmTMB v. 1.1.4 [42]. We visually examined the distributions of raw data and used the DHARMA package v. 0.4.5 [43] to aid in model diagnostics. Each model included threat duration and fighting duration as dependent measures, and genotype, day, and trial identification (ID) as random effects. See the electronic supplementary material for further details.

Broad sense genetic variance ($V_G = 2 * V_{DGRP}$), genetic correlations (r_G), and heritabilities (H^2) were extracted and computed from estimates directly from glmmTMB. We multiplied the among-DGRP hybrid variance component by 2 to account for the shared maternal line of the DGRP hybrids. We reported broad sense heritabilities because our protocol, in which we crossed males from all lines with females from a single line, did not allow us to exclude non-additive genetic effects. H^2 was estimated as $V_G / (V_G + V_E) = 2\sigma_1^2 / (2\sigma_1^2 + \sigma_e^2)$, where V_G is genetic variance, V_E is environmental variance, σ_1^2 is the among-DGRP hybrid variance component, and σ_e^2 is the error variance [44–46]. Trial level variances were used as a proxy for residual variances. Estimating quantitative genetic parameters for complex generalized linear mixed models (with respect to the scale of the latent variable versus the data scale) remains challenging for situations such as semi-continuous, ‘zero-inflated’ positive-valued data, as observed here. As such we only include these estimates on the latent variable scale, which will have higher estimates of H^2 relative to the original data scale due to the impact of exponentiation back to the response

scale [47,48]. Data and code to reproduce analyses are available at: <https://doi.org/10.6084/m9.figshare.21565722.v1> [49].

3. Results

In the experiments with (figure 1a) and without (figure 1b) females, there were relatively high genetic correlations (r_G) between wing threat and fighting ($r_G = 0.80$, 95% CI: [0.15, 0.93] and $r_G = 0.74$ [0.25, 0.89] respectively). For both data sets, we compared full model fits to their corresponding models constrained with $r_G = 0$. These comparisons are consistent with the unrestricted models being preferred for both experiments (females present: LR = 8.75, d.f. = 1, $p = 0.003$; females absent: LR = 14.2, d.f. = 1, $p = 0.0002$). Broad sense genetic variances and heritabilities for both threat and fighting were moderate (table 1).

While males spent much less time threatening than fighting when females were present (threat/fighting ratio = 0.27, with 95% CIs of 0.19–0.39, $t = -7.7$, $p < 0.0001$), they spent similar times on threat and fighting when females were absent (threat/fighting ratio = 1.18, with 95% CIs of 0.89–1.55, $t = 1.2$, $p = 0.23$, figure 1).

4. Discussion

Our experiments indicated relatively high genetic correlations between levels of threat and fighting (figure 1). The fact that threat and fighting are highly positively correlated agrees with theoretical predictions that signallers should modulate their threat in relation to their willingness to escalate into combat. Failure to do so can result in the costly defeat of weak individuals that signal strength [8–10]. Although fruit flies do not possess weapons that can inflict injuries, their fighting is nevertheless costly as it leads to reduced lifespan [50]. The theoretical predictions, however, implicitly assume that threat and fighting may vary independently. In reality, however, there are good reasons to assume that mechanistic constraints maintain at least a moderate positive genetic correlation between threat and fighting. Genetic correlations, even between apparently unrelated behaviours, are prevalent [20,22,51,52]. In the case of aggression, one can readily envision that the same genetic networks, neural networks and endocrine mechanisms modulate both threat and fighting. Finally, while we focused on the durations of threat and fighting, it is possible that other features, such as the tendency to escalate after threat, or actual ability to win a fight following a threat may be more informative.

The values of genetic correlations between threat and fighting in our experiments (0.74 and 0.80) were somewhat higher than the average of about 0.6 reported for other behaviours [20]. While such high genetic correlations may pose a

constraint on the independent evolution of threat and aggression, previous studies indicate that high genetic correlations still allow for relatively rapid evolutionary change in one of the correlated traits [19,53,54]. Intriguingly, a fruit fly study suggested that one of the five serotonin receptors they possess, 5HT_{1a}, modulates wing threat, while another serotonin receptor, 5HT₂, modulates fighting [55]. Overall though, we still do not understand how the few neurotransmitters that modulate aggression and their multiple receptors orchestrate threat and fighting [56]. Nevertheless, our results, along with the rapidly increasing mechanistic knowledge about fruit fly aggression, open up exciting avenues for future research. Specifically, given the genetic variation in the threat and fighting, one can artificially select for lineages of flies that either display few aggressive signals but eagerness to fight or vice versa. Then research on the evolved lineages can assess both the genetic and neurobiological changes in such flies and their functional effects on social interactions. All the protocols and tools for such research are currently available [28,57–59].

Males showed a much lower ratio of threat to fighting in the presence than in the absence of females (figure 1). It seems that, despite the high genetic correlation, males can adjust the ratio of threat to fighting in response to relevant features of their social environment. Males spent some time courting females and less time in both threat and aggression in the presence of

the recently mated females (panels (a) versus (b) in figure 1). While we cannot explain the difference in threat to fighting ratio in the females present and females absent experiments, audience effects on behaviour have been documented in a variety of species including fruit flies [26,34,38,60,61].

In sum, we documented a high positive correlation between aggressive signals and actual fighting. This opens up exciting opportunities for further investigations on the mechanistic and functional bases underlying the association between aggressive signals and fighting in a leading model system.

Data accessibility. Data and code to reproduce the analyses are available in the electronic supplementary material [49].

Authors' contributions. C.M.B.: conceptualization, investigation, writing—original draft, writing—review and editing; I.S.: investigation, writing—review and editing; I.D.: software, visualization, writing—original draft, writing—review and editing; R.D.: conceptualization, methodology, project administration, supervision, visualization, writing—original draft, writing—review and editing.

All authors gave final approval for publication and agreed to be held accountable for the work performed therein.

Conflict of interest declaration. We declare we have no competing interests.

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References

- Archer J. 1988 *The behavioural biology of aggression*. Cambridge, UK: Cambridge University Press.
- Huntingford F, Turner AK. 1987 *Animal conflict*. London, UK: Chapman and Hall.
- Embets Z, Somjee U, Wiens JJ. 2021 Damage from intraspecific combat is costly. *Behav. Ecol.* **32**, 1240–1245. (doi:10.1093/beheco/abab090)
- Tinbergen N. 1953 *Social behaviour in animals: with special reference to vertebrates*. London, UK: Methuen & Co.
- Crane J. 1966 Combat, display and ritualization in fiddler crabs (*Ocypodidae*, genus *Uca*). *Philos. Trans. R. Soc. Lond., B* **251**, 459–472.
- Clutton-Brock TH, Albon SD. 1979 The roaring of red deer and the evolution of honest advertisement. *Behaviour* **69**, 145–170. (doi:10.1163/156853979X00449)
- Maynard Smith J. 1974 The theory of games and the evolution of animal conflicts. *J. Theor. Biol.* **47**, 209–221. (doi:10.1016/0022-5193(74)90110-6)
- Enquist M. 1985 Communication during aggressive interactions with particular reference to variation in choice of behaviour. *Anim. Behav.* **33**, 1152–1161. (doi:10.1016/S0003-3472(85)80175-5)
- van Staaden MJ, Searcy WA, Hanlon RT. 2011 Signaling aggression. *Adv. Genet.* **75**, 23–49. (doi:10.1016/B978-0-12-380858-5.00008-3)
- Searcy WA, Nowicki S. 2005 *The evolution of animal communication: reliability and deception In signaling systems*. Princeton, NJ: Princeton University Press.
- Caryl PG. 1979 Communication by agonistic displays: what can games theory contribute to ethology? *Behaviour* **68**, 136–169. (doi:10.1163/156853979X00287)
- Baker TM, Wilson DR, Mennill DJ. 2012 Vocal signals predict attack during aggressive interactions in black-capped chickadees. *Anim. Behav.* **84**, 965–974. (doi:10.1016/j.anbehav.2012.07.022)
- Laidre ME. 2009 How often do animals lie about their intentions? An experimental test. *Am. Nat.* **173**, 337–346. (doi:10.1086/596530)
- Ballentine B, Searcy WA, Nowicki S. 2008 Reliable aggressive signalling in swamp sparrows. *Anim. Behav.* **75**, 693–703. (doi:10.1016/j.anbehav.2007.07.025)
- Searcy WA, Akçay C, Nowicki S, Beecher MD. 2014 Aggressive signaling in song sparrows and other songbirds. In *Advances in the study of behavior* (eds M Naguib, L Barrett, HJ Brockmann, S Healy, JC Mitani, TJ Roper, LW Simmons), pp. 89–125. New York, NY: Academic Press.
- Hof D, Hazlett N. 2010 Low-amplitude song predicts attack in a North American wood warbler. *Anim. Behav.* **80**, 821–828. (doi:10.1016/j.anbehav.2010.07.017)
- Templeton CN, Akçay Ç, Campbell SE, Beecher MD. 2012 Soft song is a reliable signal of aggressive intent in song sparrows. *Behav. Ecol. Sociobiol.* **66**, 1503–1509. (doi:10.1007/s00265-012-1405-5)
- Lande R, Arnold SJ. 1983 The measurement of selection on correlated characters. *Evolution* **37**, 1210–1226. (doi:10.2307/2408842)
- Dochtermann NA, Dingemans NJ. 2013 Behavioral syndromes as evolutionary constraints. *Behav. Ecol.* **24**, 806–811. (doi:10.1093/beheco/art002)
- Roff DA. 1996 The evolution of genetic correlations: an analysis of patterns. *Evolution* **50**, 1392–1403. (doi:10.1111/j.1558-5646.1996.tb03913.x)
- Conner JK, Cooper IA, La Rosa RJ, Pérez SG, Royer AM. 2014 Patterns of phenotypic correlations among morphological traits across plants and animals. *Phil. Trans. R. Soc. B* **369**, 2013246. (doi:10.1098/rstb.2013.0246)
- Saltz JB, Hessel FC, Kelly MW. 2017 Trait correlations in the genomics era. *Trends Ecol. Evol.* **32**, 279–290. (doi:10.1016/j.tree.2016.12.008)
- Chen S, Lee AY, Bowens NM, Huber R, Kravitz EA. 2002 Fighting fruit flies: a model system for the study of aggression. *Proc. Natl Acad. Sci. USA* **99**, 5664–5668. (doi:10.1073/pnas.082102599)
- Dow MA, Schilcher FV. 1975 Aggression and mating success in *Drosophila melanogaster*. *Nature* **254**, 511–512. (doi:10.1038/254511a0)
- Baxter CM, Dukas R. 2017 Life history of aggression: effects of age and sexual experience on male aggression towards males and females. *Anim. Behav.* **123**, 11–20. (doi:10.1016/j.anbehav.2016.10.022)
- Baxter CM, Barnett R, Dukas R. 2015 Aggression, mate guarding, and fitness in male fruit flies. *Anim. Behav.* **109**, 235–241. (doi:10.1016/j.anbehav.2015.08.023)
- Dukas R. 2020 Natural history of social and sexual behavior in fruit flies. *Sci. Rep.* **10**, 21932. (doi:10.1038/s41598-020-79075-7)

28. Edwards AC, Rollmann SM, Morgan TJ, Mackay TF. 2006 Quantitative genomics of aggressive behavior in *Drosophila melanogaster*. *PLoS Genet.* **2**, e154. (doi:10.1371/journal.pgen.0020154)
29. Edwards AC, Ayroles JF, Stone EA, Carbone MA, Lyman RF, Mackay T. 2009 A transcriptional network associated with natural variation in *Drosophila* aggressive behavior. *Genome Biol.* **10**, R76. (doi:10.1186/gb-2009-10-7-r76)
30. Edwards AC, Zwarts L, Yamamoto A, Callaerts P, Mackay TF. 2009 Mutations in many genes affect aggressive behavior in *Drosophila melanogaster*. *BMC Biol.* **7**, 29. (doi:10.1186/1741-7007-7-29)
31. Zwarts L, Magwire MM, Carbone MA, Versteven M, Herteleer L, Anholt RRH, Callaerts P, Mackay TFC. 2011 Complex genetic architecture of *Drosophila* aggressive behavior. *Proc. Natl Acad. Sci. USA* **108**, 17 070–17 075. (doi:10.1073/pnas.1113877108)
32. Rohde PD, Gaertner B, Wards K, Sørensen P, Mackay TF. 2017 Genomic analysis of genotype by social environment interaction for *Drosophila* aggressive behavior. *Genetics* **206**, 1969–1984. (doi:10.1534/genetics.117.200642)
33. Mackay TFC *et al.* 2012 The *Drosophila melanogaster* genetic reference panel. *Nature* **482**, 173–178. (doi:10.1038/nature10811)
34. Zuberbühler K. 2008 Audience effects. *Curr. Biol.* **18**, R189–R190. (doi:10.1016/j.cub.2007.12.041)
35. Huang W *et al.* 2014 Natural variation in genome architecture among 205 *Drosophila melanogaster* Genetic Reference Panel lines. *Genome Res.* **24**, 1193–1208. (doi:10.1101/gr.171546.113)
36. Baxter C, Yan JL, Dukas R. 2019 Genetic variation in sexual aggression and the factors that determine forced-copulation success. *Anim. Behav.* **158**, 261–267. (doi:10.1016/j.anbehav.2019.09.015)
37. Green EW, O'Callaghan EK, Hansen CN, Bastianello S, Bhutani S, Vanin S, Armstrong JD, Costa R, Kyriacou CP. 2015 *Drosophila* circadian rhythms in seminatural environments: summer afternoon component is not an artifact and requires TrpA1 channels. *Proc. Natl Acad. Sci. USA* **112**, 8702–8707. (doi:10.1073/pnas.1506093112)
38. Fitzsimmons LP, Bertram SM. 2013 Playing to an audience: the social environment influences aggression and victory displays. *Biol. Lett.* **9**, 20130449. (doi:10.1098/rsbl.2013.0449)
39. Friard O, Gamba M. 2016 BORIS: a free, versatile open-source event-logging software for video/audio coding and live observations. *Methods Ecol. Evol.* **7**, 1325–1330. (doi:10.1111/2041-210X.12584)
40. R Core Team. 2020 *R: a language and environment for statistical computing*. Vienna, Austria: R Foundation for Statistical Computing. See <http://www.R-project.org>.
41. Wickham H *et al.* 2019 Welcome to the Tidyverse. *J. Open Source Softw.* **4**, 1686. (doi:10.21105/joss.01686)
42. Brooks ME, Kristensen K, van Benthem KJ, Magnusson A, Berg CW, Nielsen A, Skaug HJ, Machler M, Bolker BM. 2017 glmmTMB balances speed and flexibility among packages for zero-inflated generalized linear mixed modeling. *R. J.* **9**, 378–400. (doi:10.32614/RJ-2017-066)
43. Hartig F. 2020 DHARMA: residual diagnostics for hierarchical (multi-level/mixed) regression models. *R package version 0.3.3.0*. **5**. See <https://CRAN.R-project.org/package=DHARMA>.
44. Scott AM, Dworkin I, Dukas R. 2018 Sociability in fruit flies: genetic variation, heritability and plasticity. *Behav. Genet.* **48**, 247–258. (doi:10.1007/s10519-018-9901-7)
45. Mackay T, Hackett JB, Lyman RF, Wayne ML, Anholt R. 1996 Quantitative genetic variation of odor-guided behavior in a natural population of *Drosophila melanogaster*. *Genetics* **144**, 727–735. (doi:10.1093/genetics/144.2.727)
46. Shorter J, Couch C, Huang W, Carbone MA, Peiffer J, Anholt RRH, Mackay TFC. 2015 Genetic architecture of natural variation in *Drosophila melanogaster* aggressive behavior. *Proc. Natl Acad. Sci. USA* **112**, E3555–E3563. (doi:10.1073/pnas.1510104112)
47. de Villemereuil P, Schielzeth H, Nakagawa S, Morrissey M. 2016 General methods for evolutionary quantitative genetic inference from generalized mixed models. *Genetics* **204**, 1281–1294. (doi:10.1534/genetics.115.186536)
48. de Villemereuil P. 2018 Quantitative genetic methods depending on the nature of the phenotypic trait. *Ann. N Y Acad. Sci.* **1422**, 29–47. (doi:10.1111/nyas.13571)
49. Baxter C, Ieta S, Dworkin I, Dukas R. 2022 Data from: Genetic correlation between aggressive signals and fighting. Figshare. (doi:10.6084/m9.figshare.21565722.v1)
50. Guo X, Dukas R. 2020 The cost of aggression in an animal without weapons. *Ethology* **126**, 24–31. (doi:10.1111/eth.12956)
51. Riechert SE, Hedrick AV. 1993 A test for correlations among fitness-linked behavioural traits in the spider *Agelenopsis aperta* (Araneae, Agelenidae). *Anim. Behav.* **46**, 669–675. (doi:10.1006/anbe.1993.1243)
52. Lynch M, Walsh B. 1998 *Genetics and analysis of quantitative traits*. Sunderland, MA: Sinauer.
53. Conner JK, Karoly K, Stewart C, Koelling VA, Sahli HF, Shaw FH. 2011 Rapid independent trait evolution despite a strong pleiotropic genetic correlation. *Am. Nat.* **178**, 429–441. (doi:10.1086/661907)
54. Beldade P, Koops K, Brakefield PM. 2002 Modularity, individuality, and evo-devo in butterfly wings. *Proc. Natl Acad. Sci. USA* **99**, 14 262–14 267. (doi:10.1073/pnas.222236199)
55. Johnson O, Becnel J, Nichols CD. 2009 Serotonin 5-HT₂ and 5-HT_{1A}-like receptors differentially modulate aggressive behaviors in *Drosophila melanogaster*. *Neuroscience* **158**, 1292–1300. (doi:10.1016/j.neuroscience.2008.10.055)
56. Asahina K. 2017 Neuromodulation and strategic action choice in *Drosophila* aggression. *Annu. Rev. Neurosci.* **40**, 51–75. (doi:10.1146/annurev-neuro-072116-031240)
57. Dierick HA, Greenspan RJ. 2006 Molecular analysis of flies selected for aggressive behavior. *Nat. Genet.* **38**, 1023–1031. (doi:10.1038/ng1864)
58. Penn JKM, Zito MF, Kravitz EA. 2010 A single social defeat reduces aggression in a highly aggressive strain of *Drosophila*. *Proc. Natl Acad. Sci. USA* **107**, 12 682–12 686. (doi:10.1073/pnas.1007016107)
59. Dukas R, Yan JL, Scott AM, Sivaratnam S, Baxter C. 2020 Artificial selection on sexual aggression: correlated traits and possible trade-offs. *Evolution* **74**, 1112–1123. (doi:10.1111/evo.13993)
60. Marler P, Dufty A, Pickert R. 1986 Vocal communication in the domestic chicken: II. Is a sender sensitive to the presence and nature of a receiver? *Anim. Behav.* **34**, 194–198. (doi:10.1016/0003-3472(86)90023-0)
61. Zajonc RB. 1965 Social facilitation. *Science* **149**, 269–274. (doi:10.1126/science.149.3681.269)